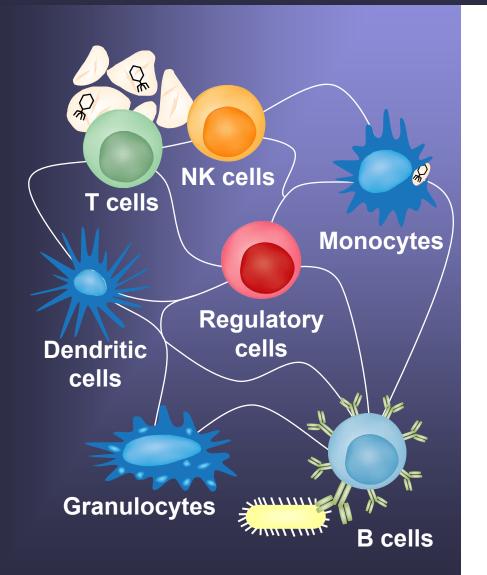


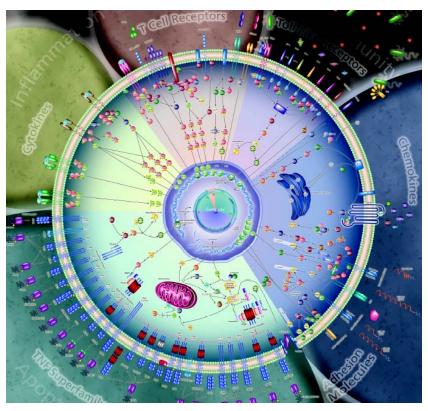
Analyzing Protein Phosphorylation Pathways in Heterogeneous Samples

Erika O'Donnell Cell Signaling Research BD Biosciences, San Diego, CA

December 1, 2011

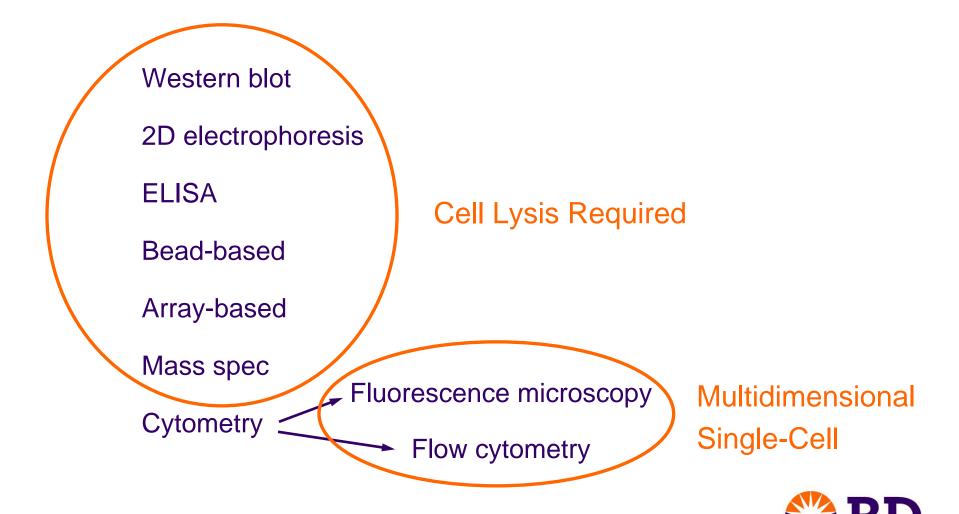
A Critical Role for Cell Signaling in Communication within the Immune System







Many Tools Are Available Today for Studying Signal Transduction

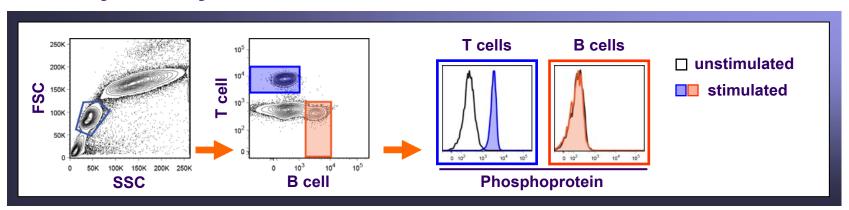


Many Tools Are Available Today for Studying Signal Transduction

Fluorescence Microscopy



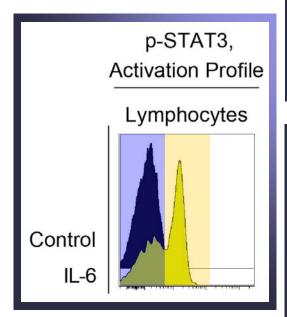
Flow Cytometry



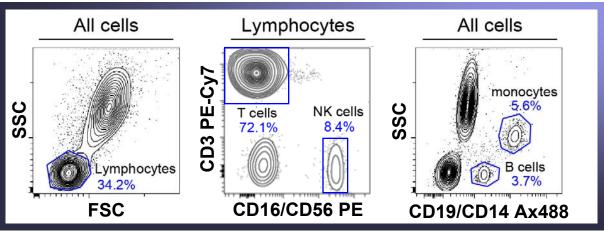


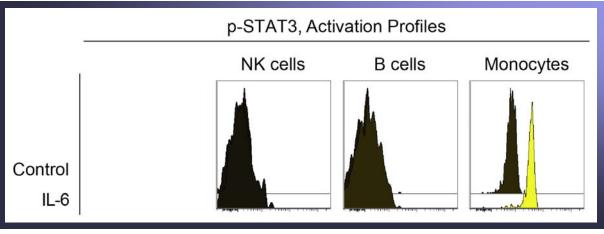
Monocyte / NK Cell Activation Kit: Signaling Responses in Human Leucocyte Subsets

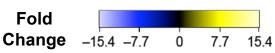
Human whole blood stimulated with the pro-inflammatory cytokine IL-6



Which lymphocytes are responding to IL-6?



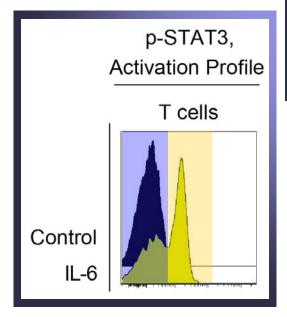




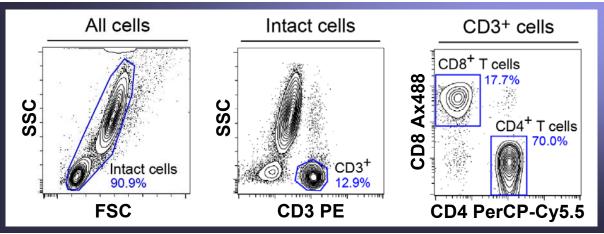


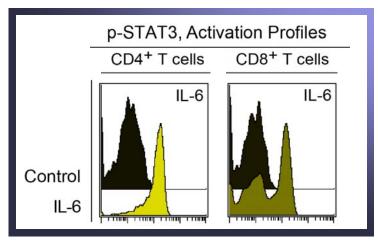
T Cell Activation Kit: Measuring Signaling Responses in CD4 and CD8 T Cells

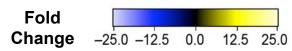
Human whole blood stimulated with the pro-inflammatory cytokine IL-6



Which T cells are responding to IL-6?









Standard Protocol for Analyzing Protein Phosphorylation by Flow Cytometry



Step 1
Stimulate cells
(optional) and
fix to preserve
phosphorylation
states



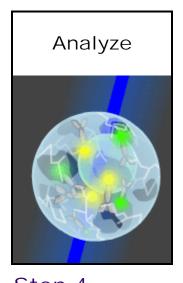
Step 2

Permeabilize
cells to allow
antibody access
to cytoplasm and
nucleus

If working with whole blood, spleen, or other erythrocyte-containing samples, RBC lysis can be performed during fixation using BD Phosflow™ Lyse/Fix Buffer



Step 3
Stain cells with
fluorescently
conjugated
antibodies against
phosphoepitopes,
surface markers,
and other targets
of interest



Step 4

Analyze cells on a properly set up flow cytometer



Elements Required for Successful Analysis of Cell Signaling at the Single-Cell Level

- Phospho-specific antibodies validated for flow cytometry
 - Specific
 - High S/N
 - Consistent
- Optimized buffer systems for fixation and permeabilization
 - Different buffer options for different sample types and phosphoepitopes
- Strategy for identification of cell populations of interest
 - Compatibility with fixation and permeabilization buffers
 - Optimization of staining conditions

- Viable and healthy samples
 - Ex vivo stimulation to trigger phospho-signaling networks
 - Detection of altered basal phosphorylation states
- Robust results
 - Careful panel design and instrument setup
 - Consistent staining
- Data analysis tools

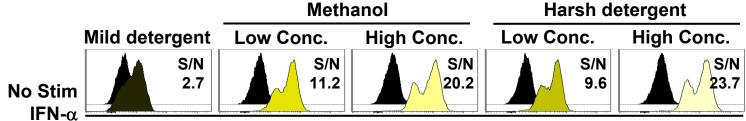


bdbiosciences.com/phosflow



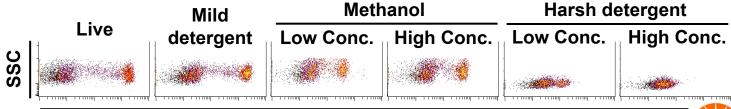
Fixation and Permeabilization: Selecting the Best Protocol

- Multiple fixation and permeabilization buffers are available
 - Appropriate buffer choice is critical for successful detection of phosphoproteins, surface markers, and other proteins of interest (eg, transcription factors, cell cycle or apoptosis proteins, etc.)
- Harsh, denaturing conditions favor detection of some phosphoproteins



Stat1 (pY701) Alexa Fluor® 647

• Fixation and permeabilization can adversely affect the detection of some surface markers, with harsh buffers causing more severe effects





Selecting Fixation and Permeabilization Buffers: Fixation

- Formaldehyde-based fixation prior to permeabilization provides optimal phosphoprotein detection and FSC/SSC resolution
 - Formaldehyde stability and concentration are critical
 - Use a source recommended in established protocols
- Sample type determines fixative choice

Sample Type	Fixative
Whole blood, spleen, or other erythrocyte-containing samples	BD Phosflow Lyse/Fix Buffer
PBMCs, cell lines, etc.	BD Cytofix™ Fixation Buffer



Selecting Fixation and Permeabilization Buffers: Permeabilization

BD Phosflow Perm Buffers

Perm/Wash Buffer I

- Mild detergent (saponin) method
- Easiest on cellsurface markers
- Adequate for detection of nuclear and cytoplasmic proteins but suboptimal for Stat pY detection

Perm Buffer II

- Mild alcohol method (low conc. methanol)
- Few cell-surface markers lost
- Good for intracellular staining

Perm Buffer III

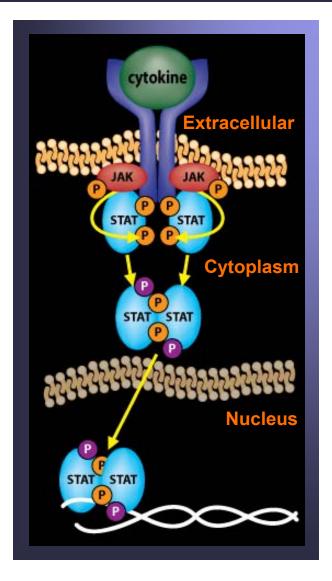
- Harsh alcohol method (high conc. methanol)
- Some cell-surface markers lost
- Best for many intracellular markers
- Most similar to Nolan lab method
- Recommended starting protocol

Perm Buffer IV

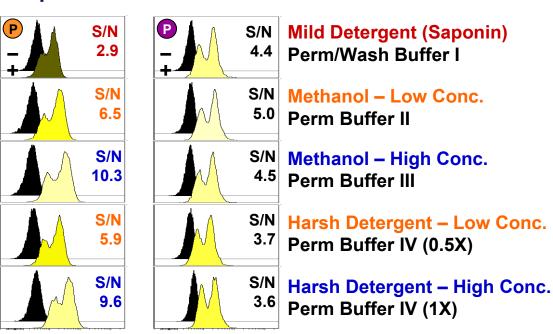
- Harsh detergent method
- Some cell-surface markers lost
- Best for Stat pY and certain other intracellular markers
- May result in greater cell loss than other buffers



Selecting Fixation and Permeabilization Buffers: Permeabilization



- Phosphoprotein detection requirements:
 - Access to cytoplasmic and/or nuclear proteins
 - Some phosphoepitopes favor harsh, denaturing permeabilization buffers

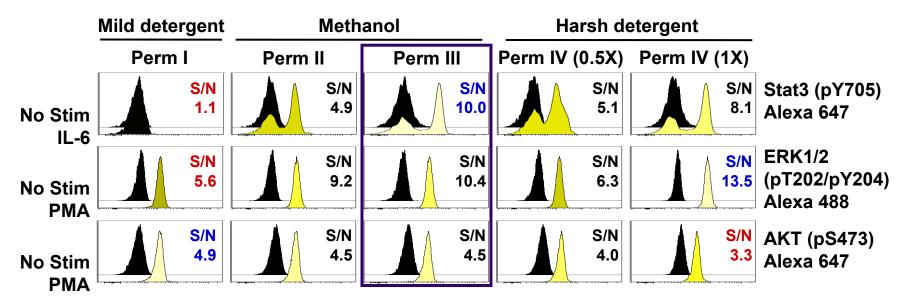


Stat1 (pY701) PE Stat1 (pS727) PE

Human PBMCs activated with IFN- α (pY701) or PMA (pS727) for 15 min and fixed with BD Cytofix



Permeabilization Buffer Selection Impacts Phosphoprotein Staining

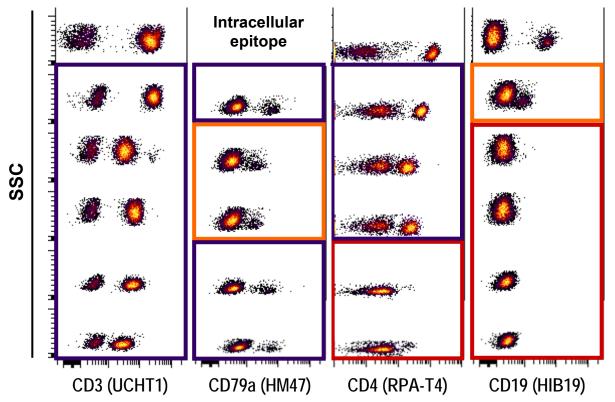


Human whole blood (Stat3) or PBMCs (ERK and AKT) activated with IL-6 or PMA for 15 min and fixed with BD Phosflow Lyse/Fix Buffer (whole blood) or BD Cytofix Buffer (PBMCs)

- Harsh permeabilization buffers provide superior staining of many, but not all, phosphoepitopes
 - High-concentration methanol (Perm III) and harsh detergent buffer (Perm IV)
- Vast majority of BD Phosflow[™] antibodies work with Perm Buffer III, although some yield superior staining with other buffers
 (eg, CREB pS133 and lκBα antibodies work best with Perm Buffer II)

Permeabilization Buffer Selection Impacts Surface Marker Resolution

- Perm/Wash Buffer I usually has the mildest effects on surface marker staining
- Different effects of methanol-based (Perm II and III) vs harsh-detergent (Perm IV) buffers on different epitopes



Live Stain
BD FACS™ Lysing Solution
post-stain

Mild Detergent (Saponin)
Perm/Wash Buffer I

Methanol – Low Conc. Perm Buffer II

Methanol – High Conc. Perm Buffer III

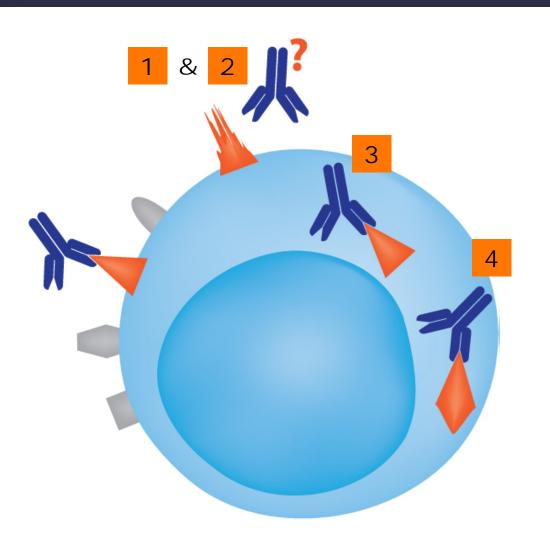
Harsh Detergent – Low Conc. Perm Buffer IV (0.5X)

Harsh Detergent – High Conc. Perm Buffer IV (1X)

Human whole blood fixed with BD Phosflow Lyse/Fix Buffer, permeabilized, and stained with PerCP-Cy™5.5–conjugated antibodies



Why is Resolution of Surface Marker Stains Reduced in Permeabilized Cells?

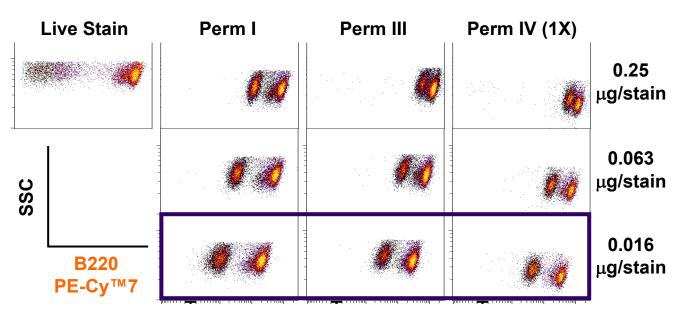


- Fixation covalently modifies surface marker epitopes, preventing antibody binding
- Harsh permeabilization buffers denature epitopes, preventing antibody binding
- Permeabilization allows antibodies access to intracellular stores of antigen
- Permeabilization opens up access to epitopes that were inaccessible during antibody screening, increasing nonspecific background



Antibody Titration Can Improve Surface Marker Resolution

Post-perm staining of anti-mouse B220 antibody is optimal at a concentration far below that used for live cell stains



BALB/c mouse spleen cells fixed with BD Phosflow Lyse/Fix Buffer, permeabilized, and stained

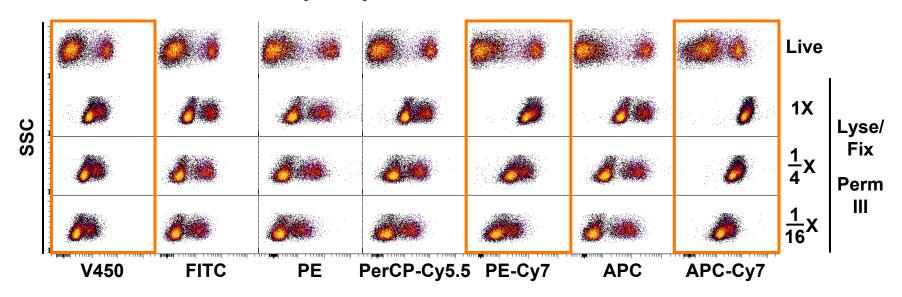


Permeabilization opens up access to epitopes that were inaccessible during antibody screening, increasing nonspecific background



Post-Perm Staining Success May Differ for Different Fluorescent Conjugates

Very high background staining and/or low signal can prevent some fluorophore conjugates of an antibody from working well in post-permeabilization stains



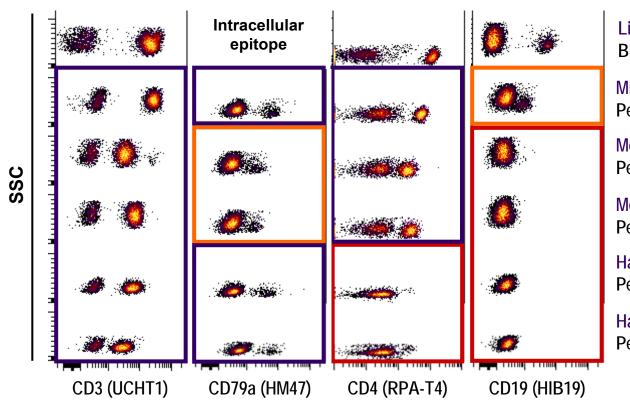
BALB/c mouse spleen cells fixed with BD Phosflow Lyse/Fix Buffer, permeabilized, and stained with various fluorophore conjugates of anti-TCRβ antibody (H57-597)



Permeabilization opens up access to epitopes that were inaccessible during antibody screening, increasing nonspecific background



Permeabilization Buffer Selection Impacts Surface Marker Resolution



Live Stain
BD FACS Lysing Solution post-stain

Mild Detergent (Saponin)
Perm/Wash Buffer I

Methanol – Low Conc. Perm Buffer II

Methanol – High Conc. Perm Buffer III

Harsh Detergent – Low Conc. Perm Buffer IV (0.5X)

Harsh Detergent – High Conc. Perm Buffer IV (1X)

Human whole blood fixed with BD Phosflow Lyse/Fix Buffer, permeabilized, and stained with PerCP-Cy5.5-conjugated antibodies



Harsh permeabilization buffers denature epitopes, preventing antibody binding



Differential Effects of Fix/Perm on Epitopes: Antibody Clone Choice is Important

Antibodies to Human Cell-Surface Markers Tested for BD Phosflow Protocols

Specificity	Clone	Fluorochrome	Protocol I Detergent method	Protocol II Mild alcohol method	Protocol III Harsh alcohol method	Protocol IV Detergent method
		APC	+	+	+	+
		APC-Cy™7	+	+	_	+
		FITC	+	+	+	+
	SK7	PE	+	+	+	+
		PE-Cy™7	+	+	+	+
		PerCP	+	+	+/-	-
		PerCP-Cy™5.5	+	+	+	+
		Alexa Fluor® 488	+	+	+	+
		Alexa Fluor® 647	+	+	+	+
		Alexa Fluor® 700		+	+/-	-
		APC	+	+	+	+
Human CD3	UCHT1	FITC	+	+	+	+
	UCHII	BD Horizon™ V450			+	+
		Pacific Blue™		+	+	+
		PE	+	+	+	+
		PE-Cy™5	+	+	+	+
		PE-Cy7	+	+	+	+
		APC	+	-	_	+/-
	HIT3a	FITC		-	_	
	niioa	PE	+	-	-	-
		PE-Cy5	+		-	-
	SP34	PE-Cy7	+		-	+/-
	5r34	PerCP	+	+	+	-



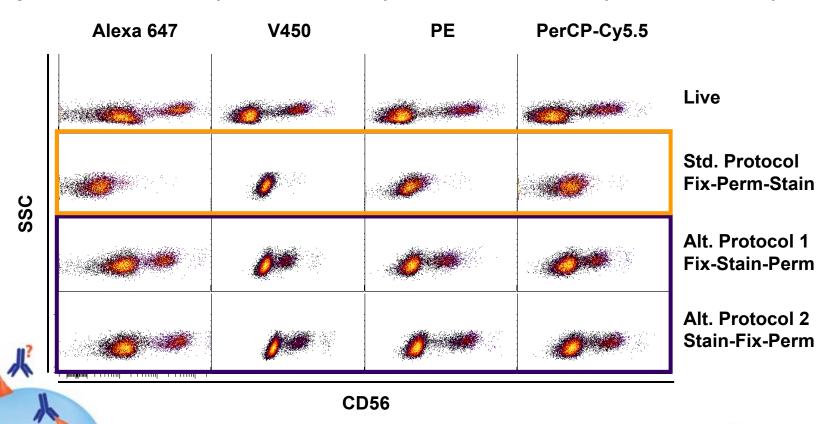
Fixation covalently modifies surface marker epitopes, preventing antibody binding

Harsh permeabilization buffers denature epitopes, preventing antibody binding



Alternative Staining Protocols Can Improve Surface Marker Resolution

Resolution of some surface markers can be improved by staining before permeabilization (Alt. Protocol 1) or before fixation (Alt. Protocol 2)

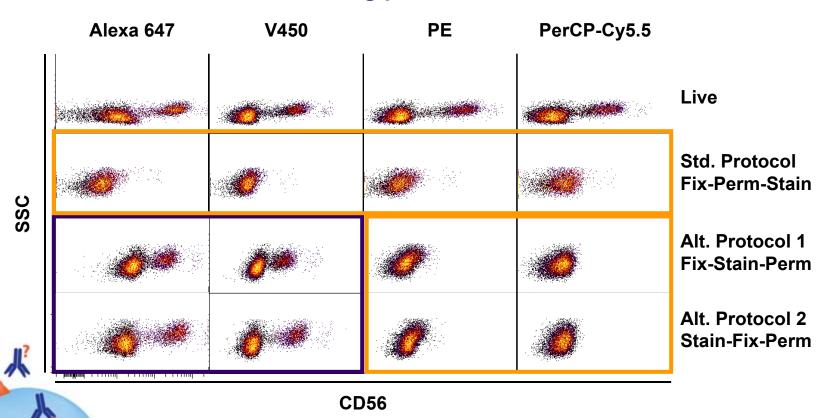


Human whole blood fixed with BD Phosflow Lyse/Fix Buffer and permeabilized with BD Phosflow Perm/Wash Buffer I



Fluorophore Choice for Alternative Staining Protocols: Perm Buffer III

Protein fluorophores are destroyed by exposure to methanol-containing permeabilization buffer

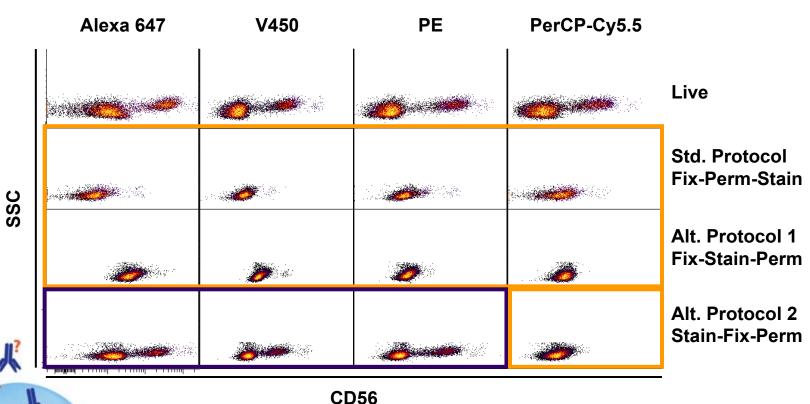


Human whole blood fixed with BD Phosflow Lyse/Fix Buffer and permeabilized with BD Phosflow Perm Buffer III



Fluorophore Choice for Alternative Staining Protocols: Perm Buffer IV

Many fluorophores are damaged by the harsh detergent-containing **BD Phosflow Perm Buffer IV, but fixation stabilizes some stains**



Human whole blood fixed with BD Phosflow Lyse/Fix Buffer and permeabilized with BD Phosflow Perm Buffer IV (1X)



BD FACSelect™ Buffer Compatibility Resource

Goal:

• To create a resource to facilitate the design of multiparameter staining panels for simultaneous analysis of intracellular and surface marker proteins

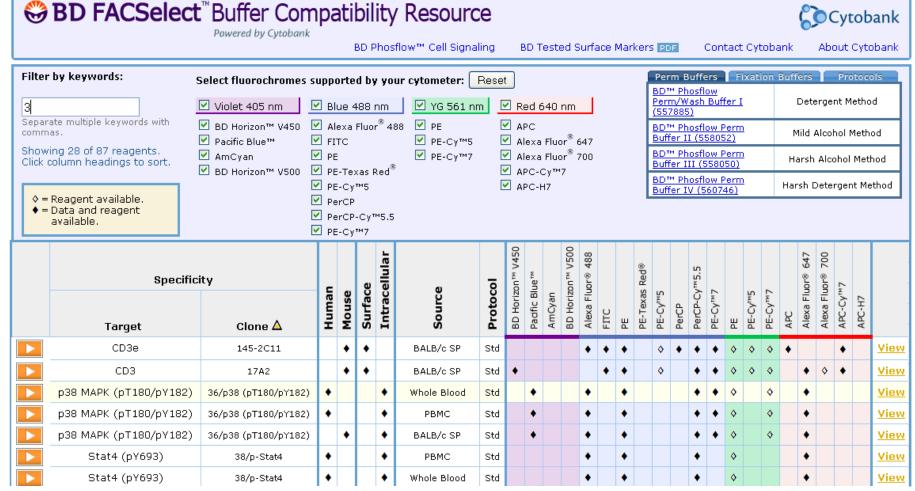
Approach:

 Generate data for key intracellular and surface marker specificities using available fluorochromes and various fixation and permeabilization protocols

Variables:

- Sample types: Human whole blood & PBMCs, murine splenocytes, & bone marrow
- Surface and intracellular specificities in all available fluorochromes
- Fixatives: BD Phosflow Lyse/Fix Buffer (human whole blood, mouse cells) or BD Cytofix Fixation Buffer (human PBMCs, cell lines)
- Permeabilization buffers: BD Phosflow Perm Buffers I–IV (IV at 1X and 0.5X conc.)
- Antibody concentrations: Three-point titrations of all surface marker antibodies
- Surface marker staining protocols: Standard Protocol (Fix-Perm-Stain), Alternative Protocol 1 (Fix-Stain-Perm), and Alternative Protocol 2 (Stain-Fix-Perm)

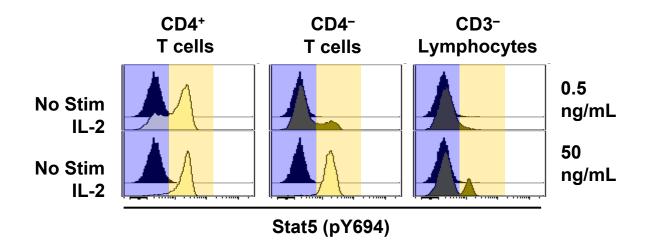
BD FACSelect™ Buffer Compatibility Resource





Designing a Multicolor Phosflow Staining Panel

Heterogeneous Threshold for IL-2 Responsiveness within Lymphocyte Subpopulations in Human Whole Blood

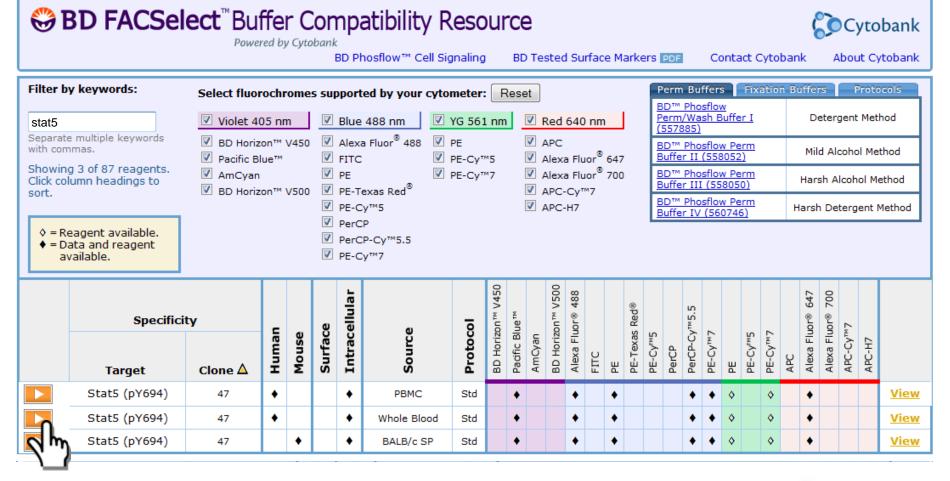


Does IL-2 responsiveness differ between naïve, effector, and memory T cells?

Need to simultaneously stain CD3, CD4, CD45RA, CD45RO, T-bet, and Stat5 (pY694) in IL-2-stimulated human whole blood cells



Step 1: Check Buffer Compatibility for Intracellular Antibodies





Stat5 (pY694) Antibody Works Well with Perm III or Perm IV (0.5x or 1x)

Stat5 (pY694) (47)



Back to FACSelect

Protein Name: Stat5 (pY694)

Clone: 47 Isotype: IgG1

Reactive species: Human, Mouse

Host species: Mouse Protocol: details

Experiment Cell Source: Human Whole Blood

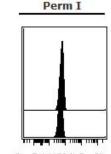
Cytometer used: FACSCantoII

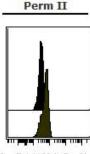
Pacific Blue™ Alexa Fluor® 488 PerCP-Cv5.5 [BD] PE-CV™7 [BD]

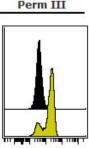
Alexa Fluor® 647 [BD]

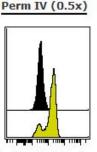
Control

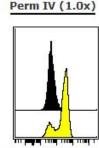
|| -2











Stat5 (pY694) PacBlu Stat5 (pY694) PacBlu Stat5 (pY694) PacBlu Stat5 (pY694) PacBlu Stat5 (pY694) PacBlu

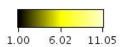
Perm L

Perm II

Perm III

Perm IV 0.5x

Perm IV 1.0x



Calculated Fold of Medians by First Row using X-Axis channel(s): Use Panel/Channel Values

Perm I Perm III Perm IV (0.5x) Perm IV (1.0x)

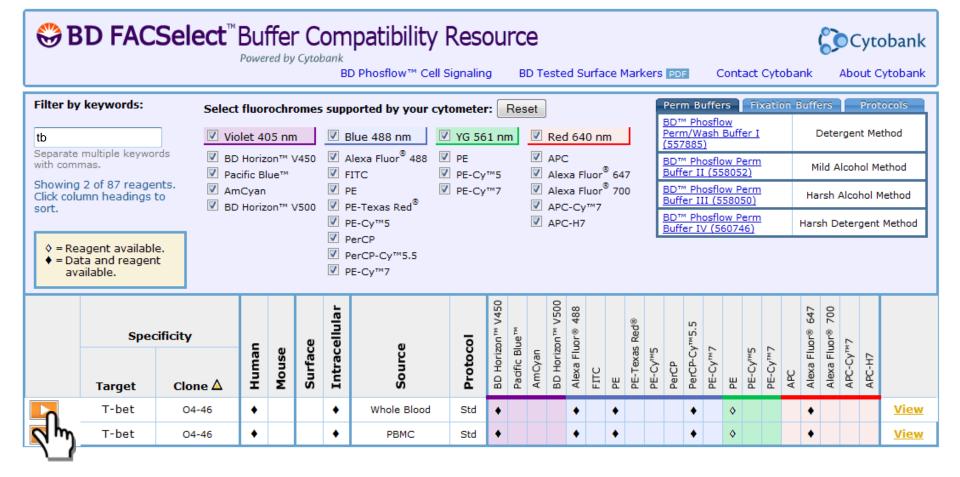
Control	1.0	1.0	1.0	1.0	1.0
IL-2	0.84	1.93	5.06	5.21	6.15







Step 1: Check Buffer Compatibility for Intracellular Antibodies





T-bet Antibody Also Works with Perm III or Perm IV (0.5x or 1x)

T-bet (04-46)



Back to FACSelect

Protein Name: T-bet Clone: 04-46

Isotype: IgG1, k

Reactive species: Human, Mouse

Host species: Mouse Protocol: details

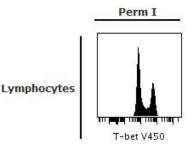
Experiment Cell Source: Human Whole Blood

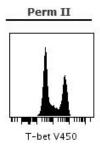
Cytometer used: FACSCantoII

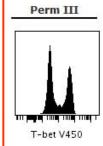
V450 [BD]

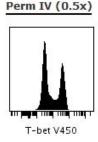
Alexa Fluor® 488 [BD] PE [BD]

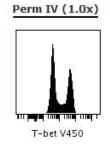
PerCP-Cv5.5 [BD] Alexa Fluor® 647 [BD]











Calculated Raw values of statistic using X-Axis channel(s): Use Panel/Channel Values

Perm I Perm II Perm IV (0.5x) Perm IV (1.0x)

2.6 3.48 3.57 3.28 3.16 Lymphocytes

View in Cytobank

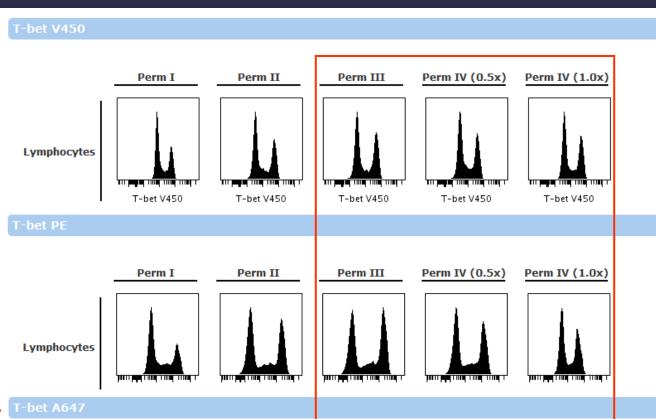
Jump to Gating Hierarchy

Back to Top

Perm I Perm II Perm III Perm IV 0.5x Perm IV 1.0x

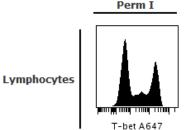


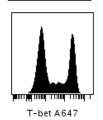
Best T-bet Resolution with Brightest Fluorophores and Perm III or IV (0.5x)



T-bet on PE or Ax647 T-bet A647

Perm I Perm II Perm IV 0.5x Perm IV 1.0x





Perm II





Perm IV (0.5x)



Perm IV (1.0x)



Step 2: Check Buffer Compatibility for **Surface Marker Antibodies**





Powered by Cytobank

BD Phosflow™ Cell Signaling

BD Tested Surface Markers PDF

Contact Cytobank

About Cytobank

Filter by keywords:

cd4

Separate multiple keywords with commas.

Showing 7 of 87 reagents. Click column headings to sort.

- = Reagent available.
- = Data and reagent available.

Select fluorochromes supported by your cytometer: Reset

- ✓ Violet 405 nm
- BD Horizon™ V450
 Alexa Fluor® 488
 PE
- ✓ Pacific Blue™
- AmCyan
- BD Horizon™ V500 PF-Texas Red®
- Blue 488 nm
- ▼ FITC
- √ PE
- - ▼ PE-Cv™5
 - ▼ PerCP
 - ✓ PerCP-Cv™5.5
 - ✓ PE-Cy™7

✓ Red 640 nm ▼ YG 561 nm

✓ APC

▼ PE-Cv^{™5}

▼ PE-Cv™7

- ✓ Alexa Fluor 647 ✓ Alexa Fluor[®] 700
- ✓ APC-Cv™7
- ✓ APC-H7

Perm Buffers Fixation	Buffers Protocols					
BD™ Phosflow Perm/Wash Buffer I (557885)	Detergent Method					
BD™ Phosflow Perm Buffer II (558052)	Mild Alcohol Method					
BD™ Phosflow Perm Buffer III (558050)	Harsh Alcohol Method					
BD™ Phosflow Perm Buffer IV (560746)	Harsh Detergent Method					

	Specific	city	_		e	ellular	ø	- 0	on™ V450	Blue™		on™ V500	Fluor® 488			. Red®			,™5.5						Fluor® 647	or® 700	47		
	Target	Clone △	Human	Mouse	Surfac	Intrac	Source	Protoc	BD Horizon	Pacific Bl	AmCyan	BD Horizon™	Alexa Flu	FITC	FE	PE-Texas	PE-Cy™5	PerCP	PerCP-Cy	PE-Cy™7	PE	PE-Cy™5	PE-Cy™7	APC	Alexa Flu	Alexa Fluor	APC-Cy"	APC-H7	
•	CD45RA	HI100	٠		٠		Whole Blood	Std	٠			\$		+	٠		٠			٠	٥	٥	٥	٠		٠		٠	<u>View</u>
	CD44	IM7		•	•		BALB/c BM	Std	+			\$		•	•		٥		•	•	٥	٥	٥	٠		٥	•		<u>View</u>
	CD45R/B220	RA3-6B2		•	•		BALB/c SP	Std	•	•		\$	•	•	\(\)	\(\)	٥	•	•	•	٥	٥	٥	•	•	٥	٠		<u>View</u>
	CD4	RM4-5		•	•		BALB/c SP	Std	٠	\$		\$	\$	•	•		•	•	•	•	٥	٥	٥	٥	•	٥			<u>View</u>
	CD4	RPA-T4	•		•		РВМС	Std	٠	٠		\$	•	•	•		•		•	•	٥	٥	٥	+	+	٠	٠	\(\)	<u>View</u>
	CD4	RPA-T4	+		٠		Whole Blood	Std	٠	٠		٥	٠	٠	٠		٠		٠	٠	٥	٥	٥	٠	+	٠	٠	♦	<u>View</u>

CD4 Clone RPA-T4 is Not Compatible with Perm Buffer IV

CD4 (RPA-T4)

Cytobank

Back to FACSelect

Protein Name: CD4 Clone: RPA-T4 Isotype: IgG1, k Reactive species: Human Host species: Mouse Protocol: details

Experiment Cell Source: Human Whole Blood

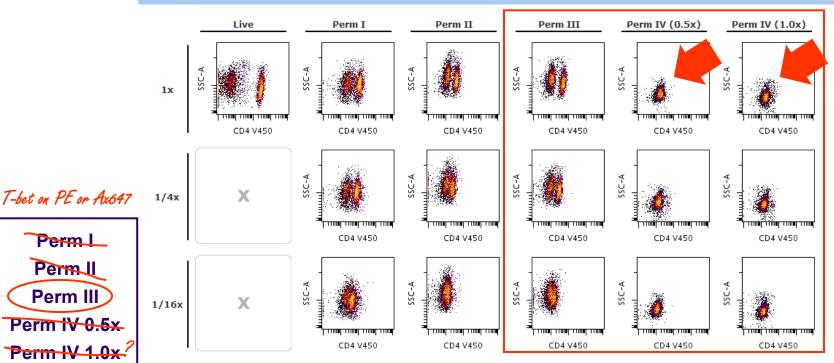
Cytometer used: LSRII

V450 [BD] Pacific Blue™ [BD] Alexa Fluor® 488 [BD] FITC [BD] PE-Cy™5 [BD] PerCP-Cv5.5 [BD] Alexa Fluor® 647 [BD] Alexa Fluor® 700 APC-CV™7

Perm L

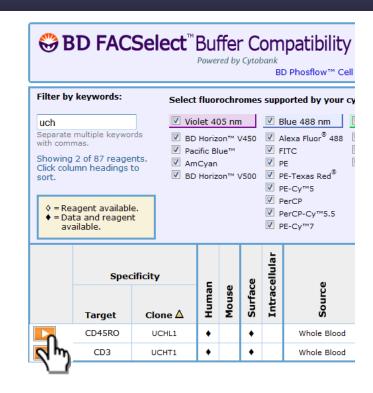
Perm II

Perm III

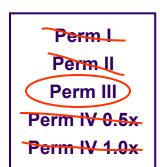


Step 3: Confirm Compatibility of All Antibodies with Selected Buffer

Specificity	Clone	Compatible with Perm Buffer III?
Stat5 (pY694)	47	✓
T-bet	O4-46	✓
CD3	UCHT1	✓
CD4	RPA-T4	✓
CD45RA	HI100	✓
CD45RO	UCHL1	✓



T-bet on PE or Ax647



Note: If some surface markers are incompatible with the chosen buffer system, alternative staining protocols may be useful. Be aware of fluorophore choice considerations.



Step 4: Select an Appropriate Conjugate for Each Antibody

Specificity	Clone	Compatible with Perm Buffer III?	Fluorophore	Optimal Concentration
Stat5 (pY694)	47	✓	Ax647	
T-bet	O4-46	✓	PE	
CD3	UCHT1	✓	Ax488	
CD4	RPA-T4	✓	PE-Cy7	
CD45RA	HI100	✓	V450	
CD45RO	UCHL1	✓	PerCP-Cy5.5	

T-bet on PE or Ax647



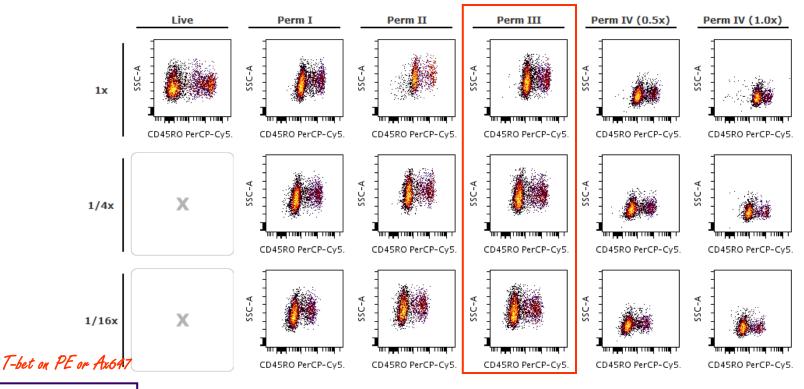
General Principles for Panel Design:

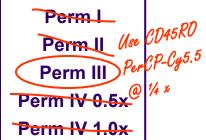
- Brightest fluorophores should be used for phospho-specific antibodies and other dim or important stains
- Try to avoid spectral overlap in channels used for phosphoprotein detection



Step 5: Identify the Optimal Concentration for Each Antibody

CD45RO PerCP-Cv5.5





Calculated Raw values of statistic using X-Axis channel(s): Use Panel/Channel Values

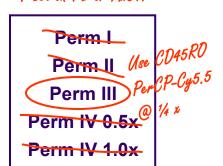
	Live	Perm I	Perm II	Perm III	Perm IV (0.5x)	Perm IV (1.0x)
1x	5.93	3.28	3.71	3.32	2.88	2.2
1/4x	Χ	3.7	3.65	4.01	3.45	2.96
1/16x	Х	3.15	3.54	3.84	3.28	3.04



Step 5: Identify the Optimal Concentration for Each Antibody

Specificity	Clone	Compatible with Perm Buffer III?	Optimal Concentration	
Stat5 (pY694)	47	✓	Ax647	Test Size
T-bet	O4-46	✓	PE	Test Size
CD3	UCHT1	✓	Ax488	1x
CD4	RPA-T4	✓	PE-Cy7	1x
CD45RA	HI100	✓	V450	1x
CD45RO	UCHL1	✓	PerCP-Cy5.5	1⁄4 X

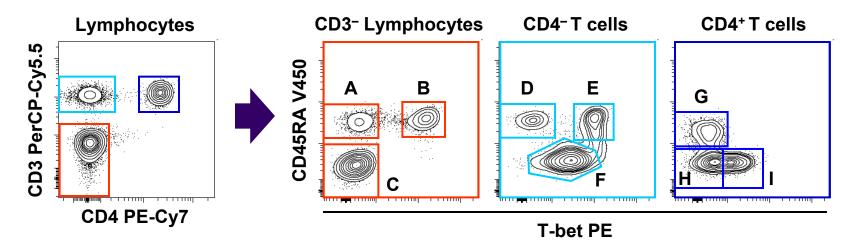
T-bet on PE or Ax647

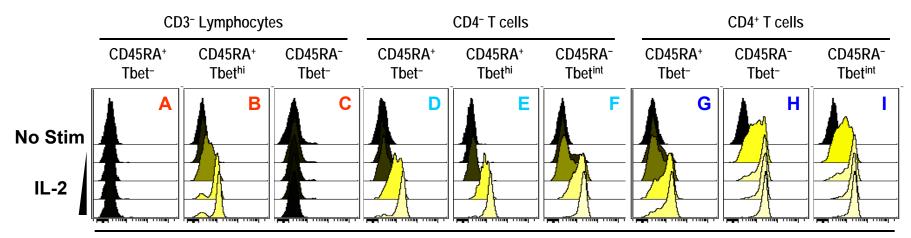


General Principles for Panel Design:

- Brightest fluorophores should be used for phosphospecific antibodies and other dim or important stains
- Try to avoid spectral overlap in channels used for phosphoprotein detection

Step 6: Stimulate, Fix, Perm, Stain, and Get Great Data





Stat5 (pY694) Ax647



Helpful Resources

BD FACSelect™ Buffer Compatibility Resource cytobank.org/facselect

BD Phosflow™ Website bdbiosciences.com/phosflow

BD FACSelect™ Multicolor Panel Designer bdbiosciences.com/paneldesigner

Manage, analyze, and share flow cytometry data over the web with Cytobank cytobank.org

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Thank You!

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